



EXPRESSION OF PAR1 IN SQUAMOUS CELL SKIN

G. Bocheva*

Department of Pharmacology and Toxicology, Medical University, Sofia

ABSTRACT:

PAR₁ is proposed to be involved in variety of invasive and metastatic cancers. Its expression correlates with the degree of invasiveness in some tissues and cancer cell lines. The aim of this study was to analyze the expression of PAR₁ in human squamous cell carcinomas with different grade of differentiation.

Immunohistochemical analysis in human SCCs and healthy skin was performed to evaluate the expression of PAR₁. The quantitative analysis demonstrated that the expression of PAR₁ was significant up-regulated in well- and moderate to poorly differentiate SCCs compared to normal skin. A significant correlation emerged between an expression of PAR₁ and differentiation of SCC. Our results could clearly show that PAR₁ expression in keratinocytes is significantly up-regulated with a decrease of tumor differentiation.

Key words: proteinase-activated receptor, squamous cell carcinoma, epidermis, skin, keratinocyte, immunohistochemistry.

INTRODUCTION

Protease-activated receptors (PARs) are a novel subfamily of seven transmembrane G protein-coupled receptors activated by serine proteinases. To date, four PARs have been cloned and characterized.

PAR₁ is proposed to be involved in variety of invasive and metastatic cancers, such as breast, colon, lung, pancreas and prostate cancer as well as malign melanoma (1-8). It was demonstrated that PAR₁ expression correlated with the degree of invasiveness in both primary breast tissues and cancer cell lines, as well as in colon cancer (5, 9, 10). This receptor is absent in normal human colon epithelial cells, but aberrantly expressed during colonic carcinogenesis (9). PAR₁ was overexpressed in tumor cells and in cells of the tumor microenvironment while low expression or absence was found in benign or normal tissues (11). Interestingly, it was shown that thrombin has a bimodal effect on PAR₁ dependent growth of melanoma, colon, and prostate carcinomas. Low concentrations of thrombin

(0,1-0,5 U/mL) enhance tumor cell growth, in contrast to higher concentrations (0,5-1 U/mL), which impair growth and induce apoptosis (7). The non-serine protease MMP-1 is also implicated in invasion and tumorigenesis of breast cancer and melanoma cells (12, 13).

Together, results from different tissues strongly support the idea that PAR₁ is involved in carcinogenesis. Although PAR₁ is highly expressed by human keratinocytes, its possible functions in the biology of malign skin tumors, however, is still unknown.

Therefore the aim of this study was to analyze the expression of PAR₁ in human squamous cell carcinomas with different grade of differentiation.

MATERIALS AND METHODS

- **Antibodies:** The primary antibody directed against human PAR₁ (clone ATAP2-mouse monoclonal) was purchased from Santa Cruz™, CA, USA) and used in 1:150 dilution.
- **Patients:** Permission for human studies was given by the Ethical Committee of the University of Münster, Germany. The study included 12 initial SCCs (10 males and 2 females, range: 42-87 years, mean age 71,0 years), 10 well differentiated SCCs (5 males and 5 females, range: 31-82

*Correspondence to: *Georgeta Bocheva*
Department of Pharmacology and Toxicology
Medical Faculty, Medical University-Sofia 1431;
"Zdrave" Str.2
e-mail: bocheva_georgeta@yahoo.com

years, mean age 62,6 years) and 9 moderate to undifferentiated SCCs (5 males and 4 females, range: 64-90 years, mean age 81,0 years). Normal skin was obtained from post-operative material (4 males and 8 females, range: 20-86 years, mean age 59,2 years). Patients did not receive topical medications within two weeks before obtaining biopsies.

- **Immunohistochemistry:** 5-6 μm paraffin sections from human skin carcinomas were heated in oven for 3h. Sections were rehydrated and heated in a steamer. Endogenous peroxidase activity was quenched with 100 mM NaN₃/0.1% H₂O₂ in PBS for 20 minutes at room temperature. Sections were further incubated with first antibody diluted in 1% BSA overnight at 4°C in a humid chamber. After rinsing with PBS, slides were incubated with second antibodies for 1 hour (DAKO En-Vision+ System Labelled Polymer-HRP™®, anti-mouse). The immunoreactivity was detected with Liquid DAB+ Substrate Chromogen System® (Dako Cytomation, Glostrup, Denmark) Sections were counterstained with Vector Hematoxylin® (Vector Laboratories, Burlingame, CA, USA, and mounted with Aguamount ® (BDH, Solms, Germany).

The staining of the tumor was scored as follows: 0: no cells positive; 1: 0-25 % positive tumor cells (weak); 2: 25-50% positive cells (moderate); 3: 50-75 % positive cells; 4: more than 75% positive tumor cells (strong).

RESULTS

Immunohistochemical analysis in human SCCs and healthy skin was performed to evaluate the expression of PAR₁. We examined 31 SCCs of different histological stages.

Immunohistochemistry revealed that in normal epidermis surrounding the tumors, PAR₁ was observed both in basal and suprabasal layers except the squamous and corneal layers. In the stroma, PAR₁ immunoreactivity could be detected in fibroblasts, endothelial cells, mast cells and neutrophils. PAR₁ expression was mainly located in the cytoplasm of the tumor cells. Immunoreactivity for PAR₁ was weak or even absent in initial SCCs (**Figure 1a**). In well and moderate to poorly differentiated SCCs, positive immunolabeling for PAR₁ was noted in 17 (89.5 %) of the tumors (**Figure 1b, c**).

In fact, quantitative analysis demonstrated that the expression of PAR₁ was significant up-regulated in well- and moderate to poorly differentiate SCCs compared to normal skin. Moreover, we found difference in PAR₁ expression between well- and poorly differentiated SCCs. A significant correlation emerged between an expression of PAR₁ and differentiation of SCC. Our results could clearly show that PAR₁ expression in keratinocytes is significantly up-regulated with a decrease of tumor differentiation. This is in line with several other observations in the literature confirming a relationship between PAR₁ expression and SCC differentiation (14).

DISCUSSION

Our knowledge about the pathogenesis of skin-derived epithelial tumors is still incomplete. Beside classical growth factors, PARs have been implicated with the control of proliferation and differentiation events in several normal and tumor tissues (5, 15-17). These effects may be at least in part mediated by activated PARs.

PAR₁ modulates keratinocyte growth and differentiation in cultured human and murine keratinocytes (18-21) and induces growth and proliferation in primary human keratinocytes (18, 20).

However, the possible function of PAR₁ in cutaneous cancerogenesis is still unknown. There is still no published data about a role of PAR₁ in non-melanoma skin cancer. Our investigation is the first to examine PAR₁ expression in squamous cell skin carcinomas (SCCs). A significant correlation emerged between an expression of PAR₁ and differentiation of SCC.

PAR₁ is well recognized as a regulator of a variety of extracellular and adhesion molecules and proposed to be involved in the invasion and metastasis of various cancers. On the other hand, PAR₁ also inhibits invasion and migration in certain tumor cells (22-24). These observations promote PAR₁ to be capable of inducing diverse intracellular signaling pathways leading to tumor progression or even tumor growth inhibition under certain circumstances. Therefore, diverse pathways of PAR₁-induced intracellular signaling pathways seem to be crucial for the invasiveness of tumor cells (25).

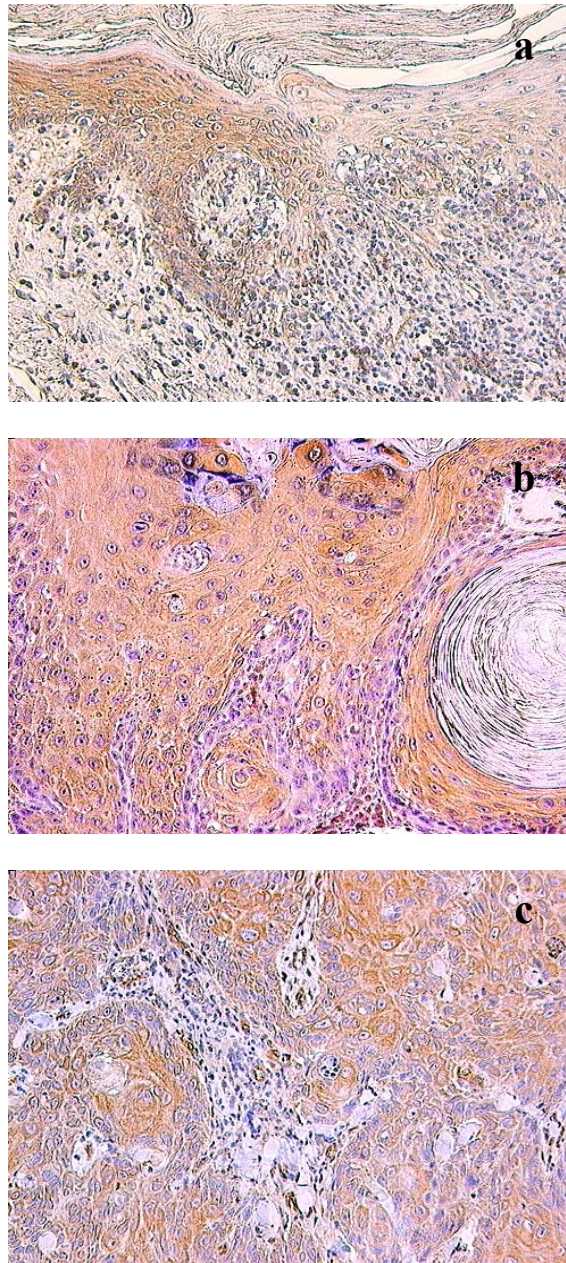


Figure 1. Expression of PAR1 in human squamocellular skin carcinomas: a) initial SCC; b) well differentiated SCC; c) moderate differentiated SCC.

REFERENCES

1. Booden M. A., Eckert LB, Der CJ, Trejo J, Persistent signaling by dysregulated thrombin receptor trafficking promotes breast carcinoma cell invasion. *Mol Cell Biol*, 24(5): 1990-9, 2004.
2. Nierodzik M. L., Chen K., Takeshita K., Li J. J., Huang Y. Q., Feng X. S., D'Andrea M. R. et al., Protease-activated receptor 1 (PAR-1) is required and rate-limiting for thrombin-enhanced experimental pulmonary metastasis. *Blood*, 92(10):3694-700, 1998.
3. Fisher E. G., Wolfram R., and Mueller B. M., Tissue factor-initiated thrombin generation activates the signaling thrombin receptor on malignant melanoma cells. *Cancer Res*, 55: 1629-32, 1995.
4. Even-Ram S., Uziely B., Cohen P., Grisaru-Granovsky S., Maoz M., Ginzburg Y. et al., Thrombin receptor overexpression in malignant and physiological invasion processes. *Nat Med*, 4(8): 909-14, 1998.

5. Even-Ram S., Maoz M., Pokroy E., Reich R., Katz B. Z., Gutwein P., Altevogt P., Bar-Shavit R., Tumor cell invasion is promoted by activation of protease activated receptor-1 in cooperation with the alpha vbeta 5 integrin. *J Biol Chem*, 276(14): 10952-62, 2001.
6. Zain J., Huang Y. O., Feng X., Nierodzik M. L., Li J. J., Karparkin S., Concentration-dependent dual effect of thrombin on impaired growth/apoptosis or mitogenesis in tumor cells. *Blood*, 95(10): 3133-8, 2000.
7. Massi D., Naldini A., Ardinghi C., Carraro F., Franchi A., Paglierani M. et al., Expression of protease-activated receptors 1 and 2 in melanocytic nevi and malignant melanoma. *Hum Pathol*, 36: 676-685, 2005.
8. Darmoul D., Gratio V., Devaud H., Lehy T., Laburthe M., Aberrant expression and activation of the thrombin receptor-1 induces cell proliferation and motility in human colon cancer cells. *Am J Pathol*, 162(5): 1503-13, 2003.
9. Darmoul D., Gratio V., Devaud H., Laburthe M., Protease-activated receptor 2 in colon cancer: trypsin-induced MAPK phosphorylation and cell proliferation are mediated by epidermal growth factor receptor transactivation. *J Biol Chem*, 279: 20927-20934, 2004.
10. D'Andrea M. R., Derian C. K., Santulli R. J., Andrade-Gordon P., Differential expression of protease-activated receptors-1 and -2 in stromal fibroblasts of normal, benign, and malignant human tissues. *Am J Pathol*, 158(6): 2031-41, 2001.
11. Boire A., Covic L., Agarwal A., Jacques S., Sherifi S., Kuliopulos A., PAR1 is a matrix metalloprotease-1 receptor that promotes invasion and tumorigenesis of breast cancer cells. *Cell*, 120(3): 303-13, 2005.
12. Goerge T., Barg A., Schnaeker E. M., Poppelmann B., Shpacovitch V., Rattenholl A., Maaser C., Luger T. A., Steinhoff M., Schneider S. W., Tumor-derived matrix metalloproteinase-1 targets endothelial proteinase-activated receptor 1 promoting endothelial cell activation. *Cancer Res*, 66(15): 7766-74, 2006.
13. Zhang X., Hunt J., Landsittel D., Muller S., Adler-Storthz K., Ferris R., Shin D., and Chen Zh., Correlation of proteinase-activated receptor-1 with differentiation markers in squamous cell carcinoma of head and neck and its implication in lymph node metastasis. *Clin Cancer Res*, 10: 8451-9, 2004.
14. Nishibori M., Mori S., Takahashi H. K., Physiology and pathophysiology of proteinase-activated receptors (PARs): PAR-2-mediated proliferation of colon cancer cell. *J Pharmacol Sci*, 97(1):25-30, 2005.
15. Macfarlane S.R., Seatter M. J., Kanke T., Hunter G. D., Plevin R., Proteinase-activated receptors. *Pharmacol Rev*, 53(2):245-82, 2001.
16. Soreide K., Janssen E. A., Körner H., Baak J. P., Trypsin in colorectal cancer: molecular biological mechanisms of proliferation, invasion, and metastasis. *J Pathol*, 209(2):147-56, 2006.
17. Santulli R. J., Derian C. K., Darrow A. L., Tomko K. A., Eckardt A. J., Seiberg M., Scarborough R. M., Andrade-Gordon P., Evidence for the presence of a protease-activated receptor distinct from the thrombin receptor in human keratinocytes. *Proc Natl Acad Sci U S A*, 92(20):9151-5, 1995.
18. Derian C. K., Eckardt A. J., Andrade-Gordon P., Differential regulation of human keratinocyte growth and differentiation by a novel family of protease-activated receptors. *Cell Growth Differ*, 8(7):743-9, 1997.
19. Algermissen B., Sitzmann J., Nürnberg W., Laubscher J. C., Henz B M., Bauer F. Distribution and potential biologic function of the thrombin receptor PAR-1 on human keratinocytes. *Arch Dermatol Res*, 292(10):488-95, 2000.
20. Meyer-Hoffert U., Rogalski C., Seifert S., Schmeling G., Wingertzahn J., Proksch E., Wiedow O., Trypsin induces epidermal proliferation and inflammation in murineskin. *Exp Dermatol*, 13(4):234-41, 2004.
21. Faivre S., Regnaud K., Bruyneel E., Nguyen Q. D., Mareel M., Emami S., Gerspach C., Suppression of cellular invasion by activated G-protein subunit Galphao, Galphai1, Galphai2, and Galphai3 and sequestration of Gbetagamma. *Mol Pharmacol*, 60: 363-72, 2001.
22. Kamath L., Meydani A., Foss F., Kuliopulos A., Signaling from protease-

- activated receptor-1 inhibits migration and invasion of breast cancer cells. *Cancer Res*, 61: 5933-40, 2001.
23. Regnaud K., Nguyen Q. D., Vakaet L., Bruyneel E., Launay J. M., Endo T., Mareel M., Gespach C., Emami S., G-protein α (alfa) subunit promotes cellular invasion, survival, and neuroendocrine differentiation in digestive and urogenital epithelial cells. *Oncogene*, 21: 4020-31, 2000.
24. Nguyen Q. D., Faivre S., Bruyneel E., Rivay C., Seto M., Endo T., Mareel M., Emami S., Gespach C., RhoA- and RhoD-dependent regulatory switch of Galpha subunit signaling by PAR-1 receptors in cellular invasion. *FASEB J*, 16: 565-76, 2002.