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Original article

CHANGES IN SERUM CORTISOL AND SOME INNATE IMMUNITY PARAMETERS AFTER EXHAUSTIVE EXERCISE IN MALE DOGS

P. V. DZHELEBOV, D. I. GUNDASHEVA, M. Y. ANDONOVA & E. P. SLAVOV

Department of General and Clinical Pathology, Faculty of Veterinary Medicine, Trakia University, 6000 Stara Zagora, Bulgaria

Summary

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The aim of the present study was to investigate the influence of exhaustive exercise on some innate immunity parameters and cortisol levels. Twelve male, mongrel dogs were divided into an experimental group, submitted to prolonged, strenuous exercise with exhaustion as the end-point, and a control group without any exposure to exercise. Serum cortisol levels were measured before exercise (BE), right after (0 h) and on 2^{nd} hour and 4^{th} hour after exercise. The neutrophil function (phagocytosis, phagocytic index, hydrogen peroxide production-H₂O₂) and classical pathway of complement activation (CPCA) were measured as followed: BE, right after exercise and on 2, 4, 24, 48, 72 h, and 7, 14 day after exercise. In experimental animals cortisol decreased on hour 4 after exercise (P<0.05), compared to BE level, and on hour 2, compared to controls (P<0.05). Percentage of H₂O₂ producing neutrophils in experimental animals dropped significantly on hour 4 and day 7 after exercise (P<0.05), compared to BE level and on hour 4 (P<0.01) vs control group. Percentage of phagocytising neutrophils decreased slightly on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 4 to be producing reutrophils decreased slightly on hour 48 (P<0.05), compared to BE level sound on hour 4 (P<0.05), compared to BE level sound on hour 4 to be producing reutrophils decreased slightly on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound on hour 48 (P<0.05), compared to BE level sound indicate inadequate adaptive response to exercise. Overtraining could make animals more susceptible to infection.

Key words: complement system, cortisol, dog, exercise, phagocytosis

INTRODUCTION

Recently exercise has become a field of intense scientific studies. Many of the studies concerning effects of exercise on endocrine and immune system are highly controversial. Results from such studies are often influenced by the animal species involved in the study, individual inherent factors (especially in humans), type of exercise and exercise protocol, nutrition, climate and other experimental conditions. Glucocorticoids are the most often investigated hormones during physical exercise. Some studies have found increase in cortisol levels after training in hunting dogs (Preziuso et al., 2000). Cortisol has been also reported to increase preexercise, due to anticipation of the exercise event, and after short-duration highintensity exercise in athletic dogs (Angle et al., 2009). After endurance running conditioned Alaskan sled dogs have showed increase in urinary cortisol, which is indicative of adrenocortical stimulation (Durocher et al., 2007). Other studies have found no changes in serum levels of cortisol after long distance running in female beagle dogs (Arokoski et al., 1993). Another study reports reduced circulating cortisol concentrations after training in rescue dogs (Rovira et al., 2008).

Short-duration strenuous sled pulling activity in dogs caused activation of neutrophils (Moritz et al., 2003). In a study involving humans, prolonged cycling did not cause any change in natural killer cell activity, neutrophil and monocyte phagocytosis (Scharhag et al., 2005). The oxidative burst of neutrophils has been reported by different researches either to decrease or increase after severe exercise like marathon running (Hessel et al., 2000; Chinda et al., 2003). The complement system in humans after severe exercise has been found to activate (Castell et al., 1997). There is very limited information concerning dogs, about the influence of exercise on innate immune mechanisms, such as phagocytosis and complement system.

The aim of the present study was to measure serum cortisol levels after a single bout of prolonged, strenuous exercise with exhaustion as the end-point, and to evaluate short- and long-term changes in neutrophil function (phagocytosis and oxidative burst) and classical pathway of complement activation in experimental canine model.

MATERIALS AND METHODS

Experimental animals

The experiment was approved by the Ethics Committee at the Faculty of Veterinary Medicine of Trakia University, Stara Zagora (Licence №17/12.07.2007). A total of twelve healthy male, mongrel dogs, 1.5 - 3.5 years old, were used in this study. Adaptation period continued one month. The dogs were treated against parasites with Biheldon (Cheironpharma Europe, each tablet containing praziquantel 50 mg, pyrantel pamoate 150 mg) at a dose of 1 tablet/10 kg. Also, they were treated against ectoparasites with antiparasite shampoo, Ectomin and Tapilan (Dorvet, Israel). An anti-rabies vaccine was also applied. Animals were kept in individual cages (situated indoors, providing constant room temperature) and went for walks twice a day - half an hour in the morning and another walk in the evening. Thus conditions were similar to the way of pet breeding. The animals were fed commercial canine dry food Cotagro adult (Cotècnica S.C.C.L.). Dogs were divided into two groups - experimental group (n=6) and control group (n=6). Dogs of experimental group weighed 18.67 ± 0.82 kg, and control animals -20.08 ± 3.29 kg. Only experimental animals were submitted to exercise.

Exercise protocol

Adaptation: During adaptation period only one person took care of the dogs so that they can get used to him and accept him as "the leader of the pack". In each day of the last week of adapting period, that person trained them to run after a bicycle supplied with a specially constructed bike leash for dogs. Dogs were willing to run and easily performed the exercise. The exercise had a very short duration of about 10 minutes and can not be considered as endurance training. Runs were performed off road to avoid trauma. During this week dogs were introduced to the exercise and to the environment so they were able to perform the exercise without distracting their attention.

Exhaustive exercise: Each dog from experimental group was submitted to a prolonged running at moderate intensity with exhaustion as the end-point. Dogs ran on a leash after a bicycle at an average speed of about 12 km/h or faster depending on dog's abilities. Dogs ran galloping and trotting. When fatigue emerged and they were unwilling to run they were encouraged by the leader verbally and by pulling the leash. Exercise was stopped when dogs could no longer sustain the needed intensity and refused to perform the exercise.

Explanatory remarks: Exhaustive exercise began with a 5 min warm up running. To prevent dehydration in every five kilometers of running dogs had access to water. Experiment was conducted under the conditions of moderate climate in spring and autumn, and ambient temperature did not exceed 20 °C. We preferred this exercise protocol instead of using a treadmill, because treadmill running is monotonous and unnatural for the dogs and they easily get bored. On the contrary when running off road they are eager and motivated to follow their "leader" as long as they can run. Dogs had no endurance training before conducting the experiment so they reached exhaustion easier through this exercise protocol.

Blood samples

Blood samples were collected in sterile glass tubes by punction of vena cephalica in the following dynamics – before exercise, right after exercise (0 h) and on 2, 4,

24, 48, 72 h and 7, 14 day after exercise. We used 0.2 mL heparin (50 units/mL) for each sample as anticoagulant. For measuring cortisol and CPCA blood was collected in tubes and was allowed to clot and then serum was separated. Blood samples for BE level and on the following days were taken at 8 a.m.

Blood analysis

• Nitroblue tetrazolium reduction test

Count of H_2O_2 producing neutrophils was calculated as percentage of total neutrophil count (100) on a safranin stained blood smear (Baehner & Nathan, 1968).

• Percentage of phagocytising neutrophils Percentage of phagocytising neutro-

Percentage of phagocytising neutrophils in whole blood samples was defined by the immunofluorescent method of Samnaliev (Samnaliev *et al.*, 1995). A total of 150 neutrophils are counted on the smear. The parameter is defined by the formula: % phagocytising cells = (count of phagocytising cells/150) × 100.

• Phagocytic index

Phagocytic index shows the mean number of engulfed bacteria by a single phagocytising cell: Phagocytic index = total count of engulfed bacteria / 150

• CPCA

The classical pathway complement activation was determined according to the method of Stelzner & Stein (1971).

Cortisol assay

The serum cortisol was determined by a competitive immunoassay using direct chemiluminescent technology (Bayer ACS:180[®] Automated Chemiluminescence Systems, Bayer Vital GmbH&Co. KG, Gesckäftsbereich Diagnostics, Germany).

Statistical analysis

Results are presented as means±SD. Data was submitted to Shapiro-Wilk's test (Sta-

tistica 6.0, by StatSoft), standard F-test and t-test (StatMost, version 2.5, provided by DataMost corporation). Differences were considered statistically significant at the P<0.05 level.

RESULTS

Two of the dogs completed 24 km of running, two of them ran 25 km and the other two covered a distance of 30 km. All dogs completed the exercise for about 2 hours (1h 58 min – 2h 9 min). Mean running distance and time were respectively 26.33 \pm 2.88 km and 122.83 \pm 4.58 min.

In experimental group cortisol levels after exercise decreased, as compared to BE levels. Changes were statistically significant (P<0.05) on 4th hour after exercise (Fig. 1). Comparison between experimental and control group in dynamics showed statistically significant lower cortisol levels in experimental group on 2^{nd} hour after exercise (P<0.05).

In experimental group, compared to BE levels, percentage of H_2O_2 producing neutrophils decreased after exercise, with the lowest number detected on hour 4 (P<0.05). Number of ROS producing neutrophils was also significantly lower on day 7 after exercise (P<0.05). Comparison between groups showed a marked decrease in experimental dogs as compared to controls on hour 4 (P<0.01), (Fig. 2).

Percentage of phagocytising neutrophils did not show any dramatic changes. In experimental group this parameter decreased on hour 48 after exercise (P<0.05). Comparison between groups did not show any significant differences (Table 1).

Both phagocytic index and CPCA showed a trend to increase after exercise with a maximum respectively on 24th and 48th hour, but changes were not statistically significant. Comparison between the two groups in dynamics also had no significant differences (Table 1).



Fig. 1. Serum cortisol levels (mean±SD) in experimental group (n=6) and control group (n=6); a¹ P<0.05 within experimental group, as compared to BE level; b¹ P<0.05 experimental vs. control group, compared at the same point of dynamics.

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Fig. 2. Percentage of H_2O_2 producing neutrophils (mean±SD) in experimental group (n=6) and control group (n=6); a¹ P<0.05 within experimental group, compared to BE level; b²P<0.01 experimental vs. control group, compared at the same point of dynamics.

In control group measured parameters did not show any significant differences.

DISCUSSION

Metabolic effects of cortisol provide normal glucose blood levels needed for the brain and serve to adapt the body when being put under extreme conditions. Surprisingly, the present study found a decrease in cortisol levels after experimental exhaustive running. This finding is in disagreement with most studies investigating the effect of different types of physical exercise (Desmecht et al., 1996; Goundasheva et al., 2005). Generally increase in cortisol levels depends both on intensity and duration, but correlates most positively with duration of exercise (Hyyppa, 2005). Field studies involving humans, horses and dogs participating in different races based on prolonged running report that levels of cortisol elevate after the race (Cook et al., 1987; Durocher et al., 2007;

Fergestad et al., 2016). The present study, although based on prolonged running, used untrained experimental animals that were not adapted to endurance exercise. If these untrained dogs where involved in endurance race event they would probably become non-finishers or would perform extremely bad. Presumably the present experimental exercise protocol had an overreaching effect and led to a short-term overload. The dogs also demonstrated a decrease in blood glucose levels (unpublished data). Some studies point that severe overtraining can lead to inappropriately low levels of cortisol (McKeever & Gordon, 2008). Mechanisms of decrease in cortisol levels under such conditions are not well understood. Some authors state that overreaching may be reflected by reduced responsiveness to ACTH, which is initially compensated for by an increased pituitary ACTH response, but decreased adrenal cortisol response (Wittert et al., 1996; Lehmann et al., 1997).

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Table 1. Percenta mean±SD.	ige of phagocy	ytisingng cells,	phagocytic index	ς, CPCA in exp	erimental grouf	p (n=6) and con	trol group (n=	6). Results are	presented as
Group	BE level	Right after exercise	2h	4h	24h	48h	72h	P7	14d
Percentage of phu	agocytising ce	(%) sili							
Experimental group	19.3±4.9	19.4±10.3	19.2±9.2	20.5±13	24±11.3	18±4.7 a ¹	22.4±3.4	20.2±12.6	23.5±9.2
Control group	18.7±2.4	19.2±5.5	17.5±3.4	18.8±6.8	19±7	18.5±4.7	19.3±5	18.2±3.4	20.4±7.6
Phagocytic index									
Experimental group	0.40 ± 0.13	0.32±0.16	0.39±0.15	0.47 ± 0.35	0.60±0.27	0.40 ± 0.06	0.50±0.11	0.50±0.38	0.57±0.22
Group group	0.28±0.05	0.37±0.16	0.38±0.10	0.39±0.18	0.39±0.15	0.36±0.14	0.33±0.19	0.29±0.12	0.39±0.20
CPCA (CH50)									
Experimental group	309±87.4	302±72.2	363.5±105.4	324.2±72.4	353±66	377.7±119.2	347±105.7	299.2±70.3	333.7±72.6
Control group	287.8±79.4	288±48.8	333.8±57.1	347.4±93	349.2±62.2	326±79.2	356.6±70	304±50	322±73.9

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a¹ P<0.05 within experimental group, as compared to BE level.

sometimes controversial. Endurance race

Other investigators suggest that imbalance between secretion and clearance may decrease cortisol. At lower work rates cortisol clearance increases disproportionally to secretion and plasma cortisol concentration may actually decrease (Cashmore *et al.*, 1977). Increased hepatic clearance is probably the leading reason for cortisol decrease. Changes in globulin levels may also be suspected to impact cortisol clearance. However, hypothalamus and pituitary or adrenal dysfunction after overtraining seems to be a more reasonable explanation for eventual decrease in cortisol (Barron *et al.*, 1985).

The insignificant elevation of cortisol in control dogs is probably the result of the excitement during blood sampling. Actually any kind of stressor leads to elevation in cortisol levels. So it is difficult to determine normal blood levels in animals, because restraint and puncture of veins always act as a stressor. And for that reason recently some less invasive methods for measuring cortisol have been developed (Bayazit, 2009; Bennett & Hayssen, 2010).

Alterations in immune functions after exercise, which are to some extent due to hormonal changes, have also been a matter of great interest. It is generally accepted that moderate exercise is beneficial for health. On the contrary acute bouts of exercise are thought to suppress immunity. Accordingly the present study found a transient decrease in phagocytic activity after running until exhaustion. However there is little evidence linking exert exercise to long-term health problems. Many aspects of immunity alterations after exercise have been studied. While increase in WBC count is consistent in different studies based on prolonged exercise (Nieman, 2000; Davis et al., 2008; Cywińska et al., 2010), changes in leukocyte functions are in horses led to a long-term decrease in neutrophil and monocyte oxidative burst activity (Robson et al., 2003). Total phagocytic activity of neutrophils also has been reported to decrease in humans after marathon race (Chinda et al., 2003). On the contrary swimming until exhaustion in guinea pigs led to a global stimulation of phagocytic function in peritoneal macrophages (Ortega et al., 1992). Other similar rodent studies exist (Su et al., 2001). Although leukocyte activity in blood and tissues may be different, some consistency about blood neutrophils exists - both phagocytic capacity and oxidative burst are decreased after endurance exercise. In most studies mechanisms underlying changes in innate immunity remain unclear. In vitro studies report influence of some cytokines and hormones on phagocytic activity of neutrophils, but in vivo phagocytosis is influenced by a variety of factors and the overall effect may differ. Prolonged running is associated with muscle damage (Kuipers, 1994), which sounds a reasonable factor for inflammatory and acute-phase response. It is expected that neutrophil phagocytosis will be enhanced under such conditions, but actually studies report reduced phagocytic capacity and reduced killing capacity per cell (Chinda et al., 2003; Robson et al., 2003). Such changes may be due to stress and overload during prolonged exercise (Nieman, 2000). It is also suggested that in the early stages of prolonged exercise phagocytic activity may be increased. Such statements seem to be logic, but quite simple and do not reveal any exact mechanism. According to some researchers decrease in oxidative burst of neutrophils may be the result of activation of antioxidant systems or down-regulation of

oxidases, which aims to prevent further

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muscle damage by oxide radicals during period after exert exercise (Tiidus, 1998). Some data exist that stress-induced suppression of immunity is seen even in adrenal-ectomized animals, which is due to the presence of immunosuppressive proteins in the blood of stressed animals (Tizard, 1996). However, with respect to immunity, decreased phagocytic activity is transient and is compensated by the marked neutrophilia (Chinda *et al.*, 2003).

Our investigation failed to find activation of the complement system. On the contrary, some studies concerning humans participating in marathon races report increase in some components of complement system, which is probably due to muscle damage (Castell et al., 1997). Other studies did not find changes in complement system (Risøy et al., 2003), which is in agreement with our results. Presumably the complement system is reacting mainly upon antigen stimulation, but not upon muscle inflammation due to exert training. This can not totally exclude the possibility that exert exercise induces local complement activation in muscle, leading to local inflammation (Risøy et al., 2003).

CONCLUSION

In conclusion, running at moderate intensity until exhaustion in untrained dogs led to decrease in cortisol levels and transient decrease in phagocytic activity of blood neutrophils, but did not alter CPCA. Decrease of cortisol could be a marker of inadequate adaptive response to endurance exercise and severe exercise could make animals more susceptible to infection. Working, hunting and sports dogs should be gradually introduced to exhaustive exercise.

REFERENCES

- Angle, C., J. Wakshlag, R. Gillette, T. Stokol, S. Geske, T. Adkins & C. Gregor, 2009. Hematologic, serum biochemical, and cortisol changes associated with anticipation of exercise and short duration highintensity exercise in sled dogs. *Veterinary Clinical Pathology*, **38**, 370–374.
- Arokoski, J., P. V. Miettinen, A. M. Säämänen, K. Haapanen, M. Parviainen, M. Tammi & H. J. Helminen, 1993. Effects of aerobic long distance running training (up to 40 km · day-1) of 1-year duration on blood and endocrine parameters of female beagle dogs. European Journal of Applied Physiology and Occupational Physiology, 67, 321–329.
- Baehner, R. & D. Nathan, 1968. Quantitative nitroblue tetrazolium test in chronic granulomatous disease. *New England Journal of Medicine*, 278, 971–976.
- Barron, J., T. Noakes, W. Levy, C. Smith & R. Millar, 1985. Hypothalamic dysfunction in overtrained athletes. *Journal of Clinical Endocrinology and Metabolism*, **60**, 803– 806.
- Bayazit, V., 2009. Evaluation of cortisol and stress in captive animals. *Australian Jour*nal of Basic and Applied Sciences, 3, 1022–1031.
- Bennett, A., & V. Hayssen, 2010. Measuring cortisol in hair and saliva from dogs: coat color and pigment differences. *Domestic Animal Endocrinology*, **39**, 171–180.
- Cashmore, G., C. Davies & J. Few, 1977. Relationship between increases in plasma cortisol concentration and rate of cortisol secretion during exercise in man. Journal of Endocrinology, 72, 109–110.
- Castell, L., J. Poortmans, R. Leclercq, M. Brasseur, J. Duchateau & E. Newsholme, 1997. Some aspects of the acute phase response after a marathon race, and the effects of glutamine supplementation. *European Journal of Applied Physiology and Occupational Physiology*, **75**, 47–53.

- Chinda, D., S. Nakaji, T. Umeda, T. Shimoyama, S. Kurakake, N. Okamura, T. Kumae & K. Sugawara, 2003. A competitive marathon race decreases neutrophil functions in athletes. *Luminescence*, 18, 324–329.
- Cook, N., A. Ng, G. Read, B. Harris & D. Riad-Fahmy, 1987. Salivary cortisol for monitoring adrenal activity during marathon runs. *Hormone Research*, 25, 18–23.
- Cywińska, A., Z. Wyszyńska, R. Górecka, E. Szarska, L. Witkowski, P. Dziekan, A. Winnicka & A. Schollenberger, 2010. The effect of the 162 km endurance ride on equine peripheral blood neutrophil and lymphocyte functions. *Polish Journal of Veterinary Science*, 13, 279–285.
- Davis, M., W. Davis, W. Ensign, K. Hinchcliff, T. Holbrook & K. Williamson, 2008. Effects of training and strenuous exercise on hematologic values and peripheral blood leukocyte subsets in racing sled dogs. Journal of the American Veterinary Medical Association, 232, 873–878.
- Desmecht, D., A. Linden, H. Amory, T. Art & P. Lekeux, 1996. Relationship of plasma lactate production to cortisol release following completion of different types of sporting events in horses. *Veterinary Research Communications*, 20, 371–379.
- Durocher, L., K. Hinchcliff, K. Williamson, E. McKenzie, T. Holbrook, M. Willard, C. Royer & M. Davis, 2007. Effect of strenuous exercise on urine concentrations of homovanillic acid, cortisol, and vanillylmandelic acid in sled dogs. *American Journal of Veterinary Research*, 68, 107– 111.
- Fergestad, M., T. Jahr, R. Krontveit & E. Skancke, 2016. Serum concentration of gastrin, cortisol and C-reactive protein in a group of Norwegian sled dogs during training and after endurance racing: a prospective cohort study. *Acta Veterinaria Scandinavica*, 58, 24–33.
- Goundasheva, D., I. Chenchev, R. Katsarova, T. Karadjov, I. Tsachev & G. Barzev, 2005. Changes in leucocyte and antibody

response following exercise in horses with booster vaccination against influenza and equine herpes virus 4/1. *Revue de Médecine Vétérinaire*, **156**, 527–532.

- Hessel, E., A. Haberland, M. Müller, D. Lerche & I. Schimke, 2000. Oxygen radical generation of neutrophils: A reason for oxidative stress during marathon running? *Clinica Chimica Acta*, 298, 145–156.
- Hyyppa, S., 2005. Endocrinal responses in exercising horses. *Livestock Production Science*, 92, 113–121.
- Kuipers, H., 1994. Exercise-induced muscle damage. International Journal of Sports Medicine, 15, 132–135.
- Lehmann, M., W. Lormes, A. Opitz-Gress, J. Steinacker, N. Netzer, C. Foster & U. Gastmann, 1997. Training and overtraining: An overview and experimental results in endurance sports. *Journal of sports medicine and physical fitness*, **37**, 7–17.
- Mc Keever, K. & M. Gordon, 2008. Endocrine alterations in the equine athlete. In: *Equine Exercise Physiology*, 1st edn, Saunders Elsevier, pp. 273–301.
- Moritz, A., B. Walcheck, J. Deye & D. Weiss, 2003. Effects of short-term racing activity on platelet and neutrophil activation in dogs. *American Journal of Veterinary Research*, 64, 855–859.
- Nieman, D., 2000. Exercise effects on systemic immunity. *Immunology and Cell Biology*, 78, 496–501.
- Ortega, E., M. Collazos, C. Barriga & M. De La Fuente, 1992. Stimulation of the phagocytic function in guinea pig peritoneal macrophages by physical activity stress. *European Journal of Applied Physiology and Occupational Physiology*, 64, 323–327.
- Preziuso, F., S. Preziuso & P. Buoncristiani, 2000. Lactate and cortisol in muscular exercise and training in hunting dog (English setter, bloodhound, griffon). *Annali della Facolta di Medicina Veterinaria di Pisa*, 53, 215–226.

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- Risøy, B., T. Raastad, J. Hallén, K. Lappegård, K. Bæverfjord, A. Kravdal, E. Siebke & H. Benestad, 2003. Delayed leukocytosis after hard strength and endurance exercise: aspects of regulatory mechanisms. *BMC Physiology*, **3**, 3–14.
- Robson, P., T. Alston & K. Myburgh, 2003. Prolonged suppression of the innate immune system in the horse following an 80 km endurance race. *Equine Veterinary Journal*, 35, 133–137.
- Rovira, S., A. Munoz & M. Benito, 2008. Effect of exercise on physiological, blood and endocrine parameters in search and rescue-trained dogs. *Veterinarni Medicina*, 53, 333–346.
- Samnaliev, M., K. Mladenow, T. Draskova & A. Radinov, 1995. Development and clinical assessment of some nonspecific factors of immunity. *Processing First National Congress of Immunity*, pp. 135–137.
- Scharhag, J., T. Meyer, H. Gabriel, B. Schlick, O. Faude & W. Kindermann, 2005. Does prolonged cycling of moderate intensity affect immune cell function? *British Journal* of Sports Medicine, **39(3)**, 171–177.
- Stelzner, A. & G. Stein, 1971. Moglichkiten zur hamolytischen Aktivitatsmessung von Gesamtkomplement. Theoretische Grundlagen und metodische Hinweise. Wissenschftliche Zeitschrift der Frieedrich Schiller Universitat Jena. *Mathematisch Naturwissenschaftliche Reine*, 20, 933–941.

- Su, S. H., H. I. Chen & C. J. Jen, 2001. Severe exercise enhances phagocytosis by murine bronchoalveolar macrophages. *Journal of Leukocyte Biology*, 69, 75–80.
- Tiidus, P., 1998. Radical species in inflammation and overtraining. *Canadian Journal* of *Physiology and Pharmacology*, **76**, 533–538.
- Tizard, I., 1996. Veterinary Immunology, 5th edn., W. B. Saunders Company, Philadelphia.
- Wittert, G., J. Livesey, E. Espiner & R. Donald, 1996. Adaptation of the hypothalamopituitary adrenal axis to chronic exercise stress in humans. *Medicine and Science in Sports and Exercise*, 28, 1015– 1019.

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Correspondence:

Petko V. Dzelebov Department of General and Clinical Pathology, Faculty of Veterinary Medicine, Trakia University, 6000 Stara Zagora, Bulgaria tel. +359 42 699 581, e-mail: petkovet@abv.bg