Original article

EVALUATION OF THE TRANSMISSION POTENTIAL AND INFECTIVITY OF H9N2 AVIAN INFLUENZA VIRUS IN DOGS FED INFECTED CHICKEN CARCASSES

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Summary


Despite the frequent reports of avian influenza A virus infections in dogs and cats, which have attracted the attention of veterinary practitioners and scientists in the respective fields of virology and epidemiology during the last decade, few reports are available regarding the natural exposure of dogs to low pathogenic viruses in field conditions. This may be important in the epidemiology of viruses in dogs on chicken farms. One of the possible causes of the interspecies transmission of viruses may be the consumption of infected carcasses by dogs. This hypothesis was examined by feeding dogs with experimentally H9N2 infected chicks. To this end, 8 dogs were selected and divided into two groups: test (n=5) and control (n=3). The dogs were fed commercial and standard diets during a 3-week adaptation period and from day 4 to 10 after being fed chicken carcasses. The test group was given H9N2 infected chicken carcasses, whereas the control group was fed on healthy ones from days 1 to 3 of the experiment. Clinical signs including depression, sneezing, coughing, nasal and ocular discharge, respiratory and heart rate changes, and rectal temperature, were monitored daily from day 0 to 10. Feces and nasal swabs were collected on a daily basis for virus detection via RT-PCR during the experimental period. HI tests for antibody titre measurement were also performed on each day of the experimental period. The results of the study indicated that the aforementioned clinical parameters remained normal in both the test and control groups. No signs of viruses were detected in the nasal swab and faeces of dogs in both groups. HI test results showed no considerable antibody titer against the H9N2 virus in either group. In conclusion, it seemed unlikely that the consumption of H9N2 infected chicken carcasses by dogs could be a cause of infection in dogs.

Key words: dog, HI, H9N2, influenza, interspecies transmission, PCR

INTRODUCTION

Influenza A viruses, as members of the Orthomyxoviridae family, are highly contagious pathogens that have been isolated from a wide variety of animals (Webster et al., 1992). Avian influenza A (AI) viruses currently circulating in avian species
are capable of infecting mammals, and their replication in mammals may facilitate their adaptation to humans (Hinshaw et al., 1981). Since the recognition of canine influenza A virus (equine H3N8) in the United States in 2004, the interest in the potential role of dogs in the ecology of influenza A viruses has been renewed (Crawford et al., 2005). Some other examples of interspecies transmission of influenza viruses include the successive interspecies transmissions of avian H3N2 among dogs (Song et al., 2008, 2009; Lee et al., 2009), the infection of dogs after scavenging on H5N1 positive chicken carcasses (Song et al., 2008) as well as infection with (human) H1N1 viruses (Dundon et al., 2010).

The interspecies transmission of influenza A virus is a crucial feature of its ecology and epidemiology (Webster, 1998) and is a possible threat to global human health (Korteweg & Gu, 2008). Dogs which come into close contact with humans may also play a role in the interspecies transmission of influenza viruses. Antigenic and genetic analyses of H9N2 viruses isolated during the last two decades indicate that these viruses are continuously evolving and have reassorted with other avian influenza viruses to generate multiple novel genotypes (Xu et al., 2004; Li et al., 2005; Xu et al., 2007). By 1997, H9N2 viruses had been isolated from multiple avian species throughout Asia, the Middle East, Europe and Africa (Alexander, 2000, Perk et al., 2006).

Unfortunately, H9N2 infection persists in many poultry farms in Iran (Nili & Asasi, 2003; Mosleh et al., 2009), and some farmers own dogs that are usually fed with dead chickens. In a recent study, Abbaszadeh et al. (2012) discovered a high serologic prevalence of influenza infections in a population of Iranian dogs. Moreover, Amirsalehy et al. (2010) experimentally demonstrated that H9N2 could infect dogs intranasally. However, there is no report in the literature about the natural exposure of dogs to H9N2 viruses. If dogs can indeed be infected by H9N2 in field conditions, they may be to a great extent responsible for the epidemiology of viruses among dogs or in chicken farms.

To evaluate the replication and transmission of the avian H9N2 virus in dogs, experiments were carried out to determine whether feeding dogs with H9N2 infected chicken carcasses could lead to significant infections in the dogs and whether this virus could be shed.

MATERIALS AND METHODS

H9N2 AI virus

The virus used for this study, that is, A/chicken/Iran/772/1998 (H9N2), was obtained from Razi Serum and Vaccine Research Institute, Tehran, Iran. The virus was propagated in 10-day-old embryo-technical chicken eggs and stored at a constant temperature of −70 °C. The AI virus was titrated to determine the 50% Egg Infectious Dose (EID₅₀) using the method of Reed & Muench (1937).

Virus inoculation to chickens

Forty one-day-old commercial broiler chickens were purchased and randomly allocated to 2 groups with equal numbers of chickens (n=20 each). The chickens had unrestricted access to food and water and were reared under biosecurity conditions. Prior to challenge (21 day of age), all birds were serologically tested against AI using the haemagglutination inhibition (HI) method. All 20 were negative for antibodies to H9N2 influenza virus antigens. On the 22nd day, the chickens in the...
second group were inoculated through the nares with $10^8$ EID$_{50}$/bird H9N2 AI virus, while those in the first group (the control group) received normal saline. On days 7, 14 and 20 post-inoculation (PI), five chickens from each group were randomly selected and used for serum sample collection. Oropharyngeal and cloacal swabs were collected from each chicken on the 3rd and 5th days post-challenge (pc) to evaluate H9N2 influenza virus infection by using RT-PCR. Chicken carcasses (whole body) were used to feed dogs on days 5 to 7 PI.

**Experimental design**

The experiment was conducted in compliance with the Animal Welfare Act (7 U.S.C. 2131 et seq.) and USDA Regulations and Policies. Eight healthy 1- to 2-year-old dogs were purchased and housed in different rooms under biosecurity conditions. The dogs were fed commercial and standard diets during a 3-week adaptation period. Blood samples were collected from all animals and serum tested to determine the presence of antibodies against H9N2 virus using the HI assay. In addition, nasal and faecal swabs from the dogs were used to evaluate H9N2 influenza virus infection via RT-PCR. All dogs tested negative to influenza H9 antigen in the HI and RT-PCR assays. Subsequently, the eight dogs were randomly divided into test (n=5) and control (n=3) groups. For three consecutive days, the test group was fed with H9N2 infected chicken carcasses, whereas the control group was given healthy ones during the same period. From day 4 to 10, all the dogs of both groups were fed with commercial and standard diets. The animals were monitored daily (8:00 AM to 2:00 PM) from day 0 to 10 in search of overt clinical signs of disease, including the presence of depression, sneezing, coughing, nasal and ocular discharge (serous, mucous, purulent, haemorrhagic or serosanguineous), changes in respiratory rate, fluctuations in heart rate, rectal temperature, and appetite condition (food consumption according to product table considered normal but less than that assumed as a decrease in appetite). Moreover, faecal and nasal swabs were collected daily for virus isolation during this period. HI tests were also performed on a daily basis (from day 0 to 10) and day 15 collected from dogs in order to measure antibody titre.

**Preparation of dog sera**

In the next stage of the experiment, serum samples collected from the dogs were treated at a constant temperature of 56 °C for 30 min to remove potential inhibitors. Subsequently, 150 μL of serum was mixed with 50 μL of 1% chicken red blood cells and incubated at room temperature for 30 min. The samples were then centrifuged at 1500 rpm for 10 min and used for HI assay.

**Hemagglutination inhibition (HI) assay**

The anti-viral antibody titres in the sera collected from the dogs and chickens were evaluated according to the HI assay guidelines outlined by the WHO Animal Influenza Training Manual (Anonymous, 2002). The HI assays were performed using homologous viruses.

**RT-PCR**

Virus RNA was extracted from faecal and nasal samples of the dogs as well as oropharyngeal and cloacal swabs of the chickens using an RNA extraction solution (RNX, Cinnagen Co., Tehran, Iran). The extraction process was conducted in accordance with the manufacturer’s instructions. Viral RNA was extracted from
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200 μL of the supernatants of 10% faecal suspensions.

The cDNA was synthesised via an AccuPowder®RT PreMix kit (BioNeer Corporation, South Korea) according to the manufacturer’s instruction. The primers were specific to the H9 protein gene (Lee et al., 2001). Five μL of total RNA and 20 pmol of each H9-specific primer were used for the preparation of cDNA. The AccuPower PCR PreMix kit was used to perform PCR so as to amplify a 488-bp fragment of the H9 protein gene of AI virus in a 20 μL reaction mixture containing 5 μL of cDNA and 10 pmol of each primer. The reaction mixture was then subjected to 35 cycles at temperatures of 94 °C, 53 °C and 72 °C for 1 min each, followed by a final extension at 72 °C for 10 min (Tajmanesh et al., 2006). The reaction was fixed at 4 °C until further use. Finally, the PCR products were separated in 1% agarose gel and visualised under ultraviolet light.

RESULTS

On day 4th after inoculation, most chickens showed depression and respiratory signs. On day 5, two chickens died, with the necropsy revealing tracheal congestion and casts in their syrinxes. The control chickens remained healthy during the study. All birds in the control group were negative for virus, while viral shedding in the faecal and oropharyngeal samples of the challenged birds was shown by RT-PCR. As shown on Fig. 1, antibody titre elevation against H9N2 virus, was observed from day 7 PI and reached 6 (log 2) on 20th day PI in the challenged group which also indicated chickens infection. Continuous reduction in maternal antibody titres was observed in control group throughout the study.

All dogs used in the study had normal respiratory rate, heart rates, and rectal temperatures before, during and after the consumption of infected chickens, with no significant changes being observed in these parameters. Moreover, there were no signs of coughing, sneezing, depression, or decreased appetite in the dogs of the control and test groups.

The mean ± standard deviation of the dogs’ antibody titre before the experiment was 4.1 ± 0.54. The HI test results for the test and control groups conducted during the study are shown on Fig. 2. There was no significant difference between the two groups. All samples from both the control and test groups of dogs were void of virus

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![Fig. 1. HI antibody titres (mean±SD) in chickens at 0, 7, 14 and 20 days after H9N2 inoculation days in control and challenged groups.](image-url)
during the study as confirmed by RT-PCR (Fig. 3).

**DISCUSSION**

Since 2004, there have been several reports of trans-species transmission of influenza A virus (FLUAV) infections in dogs. Dogs have been infected with equine influenza H3N8 (Crawford et al., 2005), avian influenza H3N2 (Song et al., 2008; 2009) and H5N1 (Song et al., 2008). In this study, the transmission potential and infectivity of H9N2 avian...
influenza virus in dogs fed with infected chicken carcasses was investigated.

Amirsalehi et al. (2010) induced avian H9N2 infection by means of intranasal inoculation in dogs. They showed that avian H9N2 influenza virus which has been isolated from outbreaks in broiler farms can infect dogs. Furthermore, affected animals expel the virus in faeces and nasal discharges. To the best of the researchers’ knowledge, there is no report about the infection of dogs with H9N2 viruses resulting from the consumption of infected carcasses in field conditions. In 2012, 209 serum specimens from dogs in Iran were screened, with no canine serum samples showing evidence of exposure to the 5 different influenza A viruses, including H9N2 (unpublished data). As previously stated in the results section, no virus was detected in the samples obtained from the test group dogs in our study, an observation which is in contrast to the results obtained by Amirsalehi et al. (2010). It should be mentioned that in the research of Amirsalehi et al. (2010), the animals were intranasally inoculated with 2 mL of (H9N2) avian influenza virus with a titer of 10^6.5 (EID50). Consequently, it is possible that swallowed viruses could cause positive samples in PCR tests of excrement. However, the virus was detected in the nasal swabs of dogs; furthermore, the inoculated group of animals which had come into contact with contaminated surfaces became seroconverted (Amirsalehi et al., 2010), so other explanations may exist for this phenomenon.

The key factor in influenza infection is the prevalence of sialic acid molecules in the virus-specific receptors (Ning et al., 2012). Receptors containing sialic acids with an α-2,3-linkage to the penultimate galactose (SAα2, 3-gal) are widely believed to be the receptors for the AI viruses, while human viruses prefer receptors that contain an α-2,6-linkage (SAα2, 6-gal) (Suzuki, 2005; Stevens et al., 2006). In birds (unlike humans), influenza virus could replicate in the GI (gastrointestinal) tract. Ning et al. (2012) showed that a dog’s respiratory tract (trachea and bronchus) is strongly positive for α-2,3-sialic acid-linked receptors, and some of these cells demonstrated weak-to-intermediate staining for α-2,6-sialic acid-linked influenza virus receptors. Surprisingly, in the GI tract (aside from the colon, caecum and rectum), most endothelial cells of the mucosa and glands in the lamina propria did not indicate influenza virus receptors. Based on the distribution of influenza virus receptors in the respiratory and GI tracts, it could be assumed that the occurrence and development of influenza virus infection in canines may be easier via the former compared to the latter. This can account for the differences between the results of the present study (where dogs were fed with infected chicken carcasses) and that carried out by Amirsalehi et al. (2010) (where intranasal inoculation of H9N2 virus to dogs was performed). In the present research, the absence of noticeable clinical signs and considerable antibody titer against H9N2 virus in HI tests, as well as the nonexistence of virus in the nasal swabs and faeces of dogs in the test group, may be due to the lack of α-2,3-sialic acid linked receptors in major parts of the GI tract of dogs. Yet, according to Amirsalehi et al. (2010), the presence of α-2,3-sialic acid-linked influenza virus receptors and some α-2,6-sialic acid-linked influenza virus receptors in the respiratory tract might have been responsible for the infections and positive nasal swabs. On the other hand, some influenza virus receptors are present in the sub-
mucosa layer and lamina propria of the mucosa of the GI tract of dogs (Ning et al., 2012); therefore, it is possible that viraemia (after intranasal inoculation) has resulted in the infection of the GI tract. This may be the reason for the positive samples in the PCR test of faeces in the study of Amirsalehi et al. (2010) and the negative results of the present one.

Low-grade fever and signs like nasal discharge, coughing and sneezing are common clinical symptoms of influenza infection in dogs (Payungporn et al., 2008). Similar to this study, Amirsalehi et al. (2010) found no significant relation between fever, heat rate, respiratory rate, or change in appetite and induced infection with H9N2. Holt et al. (2010) found no relation between dogs seropositive for H3N8 and respiratory discharges, coughing or body temperature.

Based on the present study, it appears that the consumption of H9N2 infected chicken carcasses by dogs cannot be considered as a means infection in dogs; in other words, feeding dogs with dead chickens in poultry farms may not pose a risk of transmission of H9N2 virus, although other factors, such as virus dosage or duration of exposure, could prove potential sources of infection and thus warrant further research.

REFERENCES


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