Bulgarian Journal of Veterinary Medicine (2012), 15, No 3, 184–190

SEROPREVALENCE AND ISOLATION OF ORNITHOBACTERIUM RHINOTRACHEALE IN BROILER FLOCKS IN MAZANDARAN PROVINCE, NORTH OF IRAN

S. SEIFI

Faculty of Veterinary Medicine, University of Mazandaran, Amol, Iran

Summary

Seifi, S., 2012. Seroprevalence and isolation of *Ornithobacterium rhinotracheale* in broiler flocks in Mazandaran province, north of Iran. *Bulg. J. Vet. Med.*, **15**, No 3, 184–190.

The aim of this study was to determine the prevalence of seropositive broiler flocks against *Ornithobacterium rhinotracheale* (ORT) and biochemical identification of this bacterium in Mazandaran province, north of Iran. Tracheal and serum samples were collected from 45 broiler flocks at farms and slaughter houses. From 450 tracheal samples, 12 (2.6%) ORT isolates were identified using biochemical tests. Of the 450 serum samples, 320 (71.1%) samples were positive for antibodies to ORT by ELISA. Results of this study indicated the presence of ORT and high prevalence of antibodies against ORT infection in north of Iran.

Key words: broilers, ELISA, Iran, Ornithobacterium rhinotracheale, seroprevalence

INTRODUCTION

Respiratory infections are among the most serious disease affecting poultry and cause heavy economic losses in the poultry industry worldwide. Various pathogens have been identified as causing respiratory disease, acting either in a primary or secondary role. Ornithobacterium rhinotracheale (ORT) is a gram-negative, pleomorphic, rod-shaped bacterium associated with respiratory distress, mortality, and decreased growth in chickens and turkeys (Vandamme et al., 1994; Chin et al., 2008). Ornithobacterium rhinotracheale is recognised as a primary or secondary etiological agent depending on strain virulence, adverse environmental elements, immune condition of the flock, and presence of other contagious agents in Europe, Africa, Middle East, Asia, Far East and South America (Bisgaard et al., 2008; Van Empel et al., 2008). Studies in some

parts of Turkey have shown different seroprevalence of ORT in broiler flocks (Turan & Ak, 2002; Ozbey et al., 2004). Refai et al., (2005) found that between 20.3% and 77.7% of serum samples from broiler flocks in Egypt were positive for ORT antibodies when using two different ELISA kits. Chansiripornchai et al., (2007) randomly examined 17 broiler farms (19 flocks) in Thailand. The seropositive flocks were 63% and the sera analysis showed that the individual 280 broiler serum antibody responses were 19.6% positive. Canal et al., (2003) collected 1550 sera related to 50 broiler flocks during the slaughter time in southern Brazil. The prevalence of ORT antibodies was 63.83%, but in each individual flock only 6.52% of the birds were positive. Diagnosis of ORT infections must be approved by isolation and identification of the bacteria and/or detection of antibodies using serological examination such as slide agglutination test and ELISA (Hafez, 1998; Erganis *et al.*, 2002; Ozbey *et al.*, 2004).

The presence of ORT in poultry farms in Iran has been confirmed over the last few years. Allymehr (2006) examined 463 serum samples from 50 broiler flocks in west Azarbaijan province, northwest of Iran. The result showed that 41 broiler flocks (82%) were positive for ORT. Ganbarpour & Salehi (2009) reported the seroprevalence of ORT in the southeast of Iran: 134 (31.9%) out of 420 serum samples or 17 (81%) out of 21 broiler flocks were positive.

The aim of this study was to detect ORT antibodies by ELISA from serum samples collected from commercially reared chicken flocks in Mazandaran province, north of Iran and to isolate and biochemically identify the ORT isolates from bacteriological samples.

MATERIALS AND METHODS

Sample collection

In this study a total of 250 tracheal and serum samples were collected from 25 commercially reared chicken flocks, with or without respiratory signs (10 trachea and serum samples per each flock). In addition trachea samples and serum samples were collected from 200 broilers (from 20 flocks) on the slaughter line of 4 abattoirs located in Mazandaran province, north of Iran. In the present study, the age of sampled broiler flocks was between 18 and 52 days. The samples were taken from broiler strains including Ross, Cobb, Arbor Acres and Arian, but Ross was the dominant strain. Population density differed from 5,500 to 13,000 in each flock.

BJVM, 15, No 3

Bacteriological examinations

Tracheal swabs were streaked on Mac-Conkey and sheep Blood agar with 10 µg/ml gentamicin. The plates were incubated at 37°C under aerobic conditions as well as at 37°C under microaerobic conditions in a candle jar for 2-3 days (Chin et al., 2008). Colonies, which were circular and small (1-3 mm in diameter), opaque to gravish and non-haemolytic were then selected (Vandamme et al., 1994). Colonies were stained by Gram's method and identified with biochemical methods as followed: catalase, nitrate reduction, H₂S production in triple sugar iron (TSI), ornithine decarboxylase, growth on Mac-Conkey, indole, urease, oxidase, nitrate, carbonhydrate fermentation tests such as glucose, fructose, galactose, maltose, lactose (Chin et al., 2008).

Serological examinations

The antibody titre was determined using the FlockChek® O. rhinotracheale antibody test kit (IDEXX Laboratories, Westbrook, Maine). The ELISA test and analysis of the results was performed according to the manufacturer's recommendations. Briefly, samples were diluted in a 1/500 ratio and the OD was measured on an ELISA microplate reader (SFRI IRE96, France) at 650 nm. Results were determined by calculating the sample to positive (S/P) ratio. Samples with S/P ratios of 0.4 or less were considered negative, and samples having S/P values higher than 0.4 (titres higher than 844) were considered positive.

Statistical analysis

Statistical analysis between ORT serum titers and the rate of isolation were performed by SPSS software and chi-square exact test. Seroprevalence and isolation of Ornithobacterium rhinotracheale in broiler flocks in Mazandaran...

RESULTS

Culture

A total of 450 tracheal swab samples were obtained from 45 broiler chicken flocks in Mazandaran province, north of Iran. ORT was isolated from 12 (2.6%) swab samples. The biochemical properties of 12 ORT isolates are shown in Table 1. All isolates showed the same biochemical characteristics with no deviation between them.

 Table 1. Biochemical characterisation of ORT isolates

Reaction	Result
Oxidase	+
Catalase	_
Urease	+
Indole	-
Lysine decarboxylase	-
Fructose	+
Lactose	+
Maltose	+
Galactose	+
Glucose	+
Nitrate reduction	-
TSI	-
Growth on MacConkey agar	_

ELISA

Of the 450 serum samples obtained from 45 broiler chicken flocks, 320 (71.1%) samples were positive for the presence of antibodies to ORT by ELISA. All of the flocks with respiratory signs were serologically positive. Details of broiler flocks with bacteriological and serological results are shown in Table 2. There was no significant difference between flock size and isolation of ORT; the age of birds was neither a significant factor in ORT isolation.

There was no significant difference between the rate of isolation and flock ORT titers (P>0.05).

DISCUSSION

Mazandaran province is located in north of Iran beside the Caspian Sea. The climate is temperate as an effect of Alborz Mountainous climate and the Caspian Sea. Being adjacent to the Caspian Sea, the climate of the province is highly humid (relative humidity is between 40 to 100%) and average temperature is 17.5 °C. It may provide optimal condition for ORT and affects the high prevalence of this bacterium. As clinical signs and post-mortem lesions of ORT infection are similar to other bacterial and virus infections, accurate diagnosis must be substantiated by direct detection or isolation of the causative bacteria and /or indirectly through the detection of antibodies using serological examination (Van Empel & Hafez, 1999; Hafez, 2002).

In this study 5 out of 25 broiler flocks showed respiratory signs such as sneezing, coughing and nasal and ocular discharge by the time of sampling and all of them were serologically positive. This finding is in agreement with result of Canal *et al.*, (2003) who found a positive correlation between the presence of respiratory signs and antibodies to ORT, although the reverse correlation was not significant. Furthermore, 9 out of 20 broiler flocks sampled at slaughterhouses were serologically positive. This finding indicates that apparently healthy flocks can be carriers of the disease.

The presence of antibodies against *O*. *rhinotracheale* could be detected by ELISA in one-day-old birds and in egg

No. of	Flocks	Age	Number of sampled chickens		Number (%) positive	
HOCKS	SIZE	(uays)	Bacteriology	Serology	Bacteriology	Serology
1	7000	32	10	10	0	5
2	6500	38	10	10	0	7
3	10800	27	10	10	0	6
4	11000	41	10	10	1	10
5	7000	34	10	10	0	7
6	8700	34	10	10	0	6
7	13000	28	10	10	1	10
8	10000	31	10	10	0	6
9	6600	18	10	10	Õ	6
10	9000	33	10	10	1	10
11	7000	41	10	10	0	10
12	6500	25	10	10	Ő	6
13	5500	27	10	10	1	10
14	12000	31	10	10	1	10
15	7500	19	10	10	0	6
16	13000	27	10	10	0 0	5
10	6500	20	10	10	0	6
18	6500	20	10	10	1	9
10	9200	38	10	10	0	8
20	8800	22	10	10	0	7
20	6000	22	10	10	0	8
21	5500	10	10	10	0	8 7
22	12000	26	10	10	1	10
23	12000	20	10	10	1	10
24	10000	33 43	10	10	0	10
25	6600	43	10	10	1	10
20	5500	32	10	10	1	9
27	11200	40	10	10	0	1 6
20	11200	50	10	10	0	07
29	12700	50	10	10	0	/
30	12/00	50	10	10	0	8
31	8000	48	10	10	0	6
32	9000	43	10	10	1	
33	8/00	4 /	10	10	0	6
34	9100	42	10	10	0	0
35	10000	55 52	10	10	1	10
36	11300	53	10	10	0	
3/	13000	46	10	10	0	6
38	7000	45	10	10	0	6
39	6500	50	10	10	1	10
40	8000	47	10	10	0	0
41	9500	55	10	10	0	1
42	12000	43	10	10	0	6
43	10500	51	10	10	0	7
44	7000	48	10	10	0	6
45	8000	49	10	10	0	6
Total			450	450	12 (2.6%)	320 (71.1%)

Table 2. Characteristics of sampled broiler flocks with bacteriologic and serologic results

BJVM, 15, No 3

yolks, as well as in birds with clinical signs, confirming that the ELISA can be useful for diagnostic purposes. Antibody titres peak at 1 to 4 weeks after a field infection, but decline rapidly thereafter, indicating that serum samples for flock screening should be taken at different ages (Van Empel & Hafez, 1999).

In this study, ORT was isolated from 2.6% of the samples. This proportion is lower than other researchers' results, such as: 9.8% in Markazi province, Iran (Ghaemmaghami et al., 2007), 8.8% in Northern and Middle Jordan (El-Sukhon et al., 2002) and 11.46% in Turkey (Turan & Ak, 2002), but is in agreement with 3.5% in south-eastern Iran (Ghanbarpour & Salehi, 2009), 2.27% in Guilan province, north of Iran (Asadpour et al., 2008), 1% in Ahvaz, South of Iran (Jamshidian & Mayahi, 2008), 1.5% in Elazig province, East of Turkey (Ozbey et al., 2004), 0.4% (Erganis et al., 2002), and 1.2% in Turkey (Turkyilmaz, 2005). ORT can normally be isolated only at an early stage of the infection and attempts to recover it at a later stage often fail. In contaminated samples ORT can be masked easily by an overgrowth of other bacteria such as E. coli, Proteus spp. or Pseudomonas spp. and therefore cannot be detected in routine investigations. Conversely, serology is useful for flock monitoring or as an aid in the diagnosis of ORT (Hafez, 2002; Chin et al., 2008).

In this study, serum samples were positive in 71.1% of samples by ELISA. These findings are higher than results recorded in Iran by Ghanbarpour & Salehi (2009) and Allymehr (2006) (31.9%, and 44.2%, respectively), but are in agreement with the results reported by Refai *et al.*, (2005) from broiler flocks in Egypt and by Turan & Ak (2002) from broiler flocks in Turkey (77.7% and 64.4%, respectively). Another study in Iran reported that 97.63% of serum samples were positive for ORT antibodies by ELISA (Keleidari *et al.*, 2008).

In a field study using a sensitive technique for confirmation, ORT was found to be involved in 70% of cases with respiratory signs in broiler chickens, while through bacteriology and/or serology only 30% of the cases could be related to ORT (Pattison et al., 2008). In the current study ORT was isolated from a low percentage of ELISA positive broiler flocks. In Brazil, ORT strains were isolated from 16% of the ELISA positive broiler flocks (Canal et al., 2005). The difference may be related to the age of the examined chickens and sample collection approach. ORT excretion and antibody response may also be affected by a number of factors such as antibiotic therapy and vaccination. The effect of antibiotic therapy on the serologic response to ORT remains unclear (Hafez, 2002; Ozbey et al., 2004).

In conclusion, the results of this study indicated for the first time the presence of ORT and high prevalence of antibodies against ORT infection in Mazandaran province. Determination of different serotypes of ORT can useful in disease prevention programme. Future studies on the current topic are therefore necessary.

ACKNOWLEDGMENTS

This work was supported by the Research Council of University of Mazandaran.

REFERENCES

Allymehr, M., 2006. Seroprevalence of Ornithobacterium rhinotracheale infection in broiler and broiler breeder chickens in west Azerbaijan province, Iran. Journal of Veterinary Medicine Series A, 53, 40–42.

- Asadpour, Y., M. H. Bozorgmehri Fard, S. A. Pourbakhsh, M. Banani & S. Charkhkar, 2008. Isolation and identification of *Ornithobacterium rhinotracheale* in broiler breeder flocks of Guilan province, north of Iran. *Pakistan Journal of Biological Science*, **11**, 1484–1491.
- Bisgaard, M., A. M. Bojesen, J. P. Christensen, P. Mark, F. M. Paul, M. B. Janet & J. A. Dennis, 2008. Infections caused by species of *Pasteurellaceae*, *Ornithobacterium* and *Riemerella*: An introduction. In: *Poultry Diseases*, 6th edn, W.B. Saunders, Edinburgh, pp. 146–148.
- Canal, C. W., J. A. Leao, S. L. S. Rocha, M. Macagnan, C. A. V. Lima-Rosaa, S. D. Oliveiraa & A. Back, 2005. Isolation and characterization of *Ornithobacterium rhinotracheale* from chickens in Brazil. *Research in Veterinary Science*, **78**, 225– 230.
- Canal, C. W., J. A. Leao, D. J. Ferreira, M. Macagnan, C. T. Pippi Salle & A. Back, 2003. Prevalence of antibodies against *Ornithobacterium rhinotracheale* in broilers and breeders in Southern Brazil. *Avian Disease*, 47, 731–737.
- Chansiripornchai, N., W. Wanasawaeng & J. Sasipreeyajan, 2007. Seroprevalence and identification of ORT from broiler and broiler breeder flocks in Thailand. *Avian Disease*, 51, 777–780.
- Chin, R. P., P. C. M. Van Empel & H. M. Hafez, 2008. Ornithobacterium rhinotracheale infection. In: Diseases of Poultry, 12th edn, Blackwell Publishing, Iowa, USA, pp. 765–774.
- El-Sukhon, S. N., A. Musa & M. Al-Attar, 2002. Studies on the bacterial etiology of airsacculitis of broilers in Northern and Middle Jordan with special reference to *Escherichia coli*, Ornithobacterium rhinotracheale and Bordetella avium. Avian Diseases, 46, 605–612.
- Erganis, O., H. H. Hadimli, K. Kav, M. Çorlu & D. Ozturk, 2002. A comparative study on detection of *Ornithobacterium rhinotracheale* antibodies in meat type tur-

keys by dot immuno binding assay, rapid agglutination test and serum agglutination test. *Avian Pathology*, **31**, 201–204.

- Ghaemmaghami, S. H., J. Vande Yousefi, H. Niroumand, A. Monsefi & S. Ahmadloo, 2007. Survey of prevalence of *Ornithobacterium rhinotracheale* in broiler farms affected with respiratory disorders in Markazi province. *Journal of Veterinary Research*, **62**, 297–300.
- Ghanbarpour, R. & M. Salehi, 2009. Seroprevalence and identification of Ornithobacterium rhinotracheale in broiler flocks in south-eastern Iran. Tropical Animal Health and Production, 41, 1679–1683.
- Hafez, H. M., 1998. Current status on the laboratory diagnosis of Ornithobacterium rhinotracheale "ORT" in poultry. Berliner und Münchener Tierärztliche Wochenschrift, 111, 143–145.
- Hafez, H. M., 2002. Diagnosis of Ornithobacterium rhinotracheale. International Journal of Poultry Science, 1, 114–118.
- Jamshidian, M. & M. Mayahi, 2008. Isolation of Ornithobacterium rhinotracheale from broilers in Ahvaz. Iranian Veterinary Journal, 4, 29–36.
- Keleidari, G. A., M. R. Basami, H. Kavoosi & F. Kordi, 2008. Seroprevalence of Ornithobacterium rhinotracheale in a selected number of broiler and a broiler breeder by the use of ELISA assay in Mashhad. In: Proceedings of the 4th National Symposium of Poultry Disease, Shahrekordm pp. 5–9.
- Ozbey, G., H. Ongor, D. T. Balik, V. Celik, A. Kilic & A. Muz, 2004. Investigations on *Ornithobacterium rhinotracheale* in broiler flocks in Elazig province located in the East of Turkey. *Veterinary Medicine* – *Czech*, **49**, 305–311.
- Pattison, M., P. F. McMullin, J. M. Bradbury & D. J. Alexander, 2008. Poultry Diseases, 6th edn, Saunders Elsevier, London, pp. 164–170.
- Refai, M., A. El-Gohary, S. A. Attia, R. A. Khalifa, 2005. Diagnosis of *Ornithobacterium rhinotracheale* infection in chickens

S. Seifi

BJVM, 15, No 3

Seroprevalence and isolation of Ornithobacterium rhinotracheale in broiler flocks in Mazandaran...

by ELISA. *Egyptian Journal of Immunol- ogy*, **12**, 87–93.

- Turan, N. & S. Ak, 2002. Investigation of the presence of Ornithobacterium rhinotracheale in chickens in Turkey and determination of the seroprevalance of the infection using the enzyme-linked immunosorbent assay. Avian Diseases, 46, 442–446.
- Turkyilmaz, S., 2005. Isolation and serotyping of Ornithobacterium rhinotracheale from poultry. Turkish Journal of Veterinary and Animal Sciences, 29, 1299–1304.
- Van Empel, P., P. Mark, F. M. Paul, M. B. Janet & J. A. Dennis, 2008. Ornithobacterium rhinotracheale. In: Poultry Diseases, 6th edn, W. B. Saunders, Edinburgh, pp. 164–171.
- Van Empel, P. C. M. & H. M. Hafez, 1999. Ornithobacterium rhinotracheale: A review. Avian Pathology, 28, 217–227.
- Vandamme, P., P. Segers, M. Vancaneyt, K. Van Hover, R. Mutters, J. Hommez, F. Dewirst, B. Paster, K. Kersters, E. Falsen, L. Devrieze, M. Bisgaard, K. H. Hinz & W. Mannheim, 1994. Description of Ornithobacterium rhinotracheale gen. nov., sp. nov., isolated from the avian respiratory

tract. International Journal of Systematic Bacteriology, 44, 24–37.

Paper received 16.02.2012; accepted for publication 21.06.2012

Correspondence:

Dr. Saeed Seifi Faculty of Veterinary Medicine, University of Mazandaran, Amol, Iran. P.O. Box: 46168-49767, tel: +981212271055, fax: +981212271054, e-mail: saeedseifi@umz.ac.ir